

Note

Synthesis of acylphosphonyl analogues of phosphatidyl-*myo*-inositol as inhibitors of phosphatidylinositol-specific phospholipase C

Maria A. Alisi ^a, Mario Brufani ^a, Luigi Filocamo ^{a,*},
Luciano Cellai ^b, M. Adelaide Iannelli ^b, Maria C. Cesta ^c,
Sperandina Lappa ^c

^a *Dipartimento di Scienze Biochimiche "A. Rossi Fanelli", Università "La Sapienza", Via degli Apuli 9, 00185 Roma, Italy*

^b *Istituto di Strutturistica Chimica "G. Giacomello", C.N.R., c.p. 10, 00016 Monterotondo stazione, Roma, Italy*

^c *Mediolanum Farmaceutici S.p.a., Via S.G. Cottolengo 31, 20143 Milano, Italy*

Received 25 March 1994; accepted in final form 17 June 1994

Keywords: Synthesis; Acylphosphonyl analogues; Phosphatidyl-*myo*-inositol; Phospholipase

It is now well-known that *myo*-inositol 1,4,5-trisphosphate and diacylglycerol are released upon phosphatidylinositol-specific phospholipase C (PI-PLC) catalysed cleavage of phosphatidylinositol 4,5-bisphosphate after agonist stimulation of several cell surface receptors [1]. The PI-PLC is an ubiquitous enzyme and it is involved in various cellular functions such as signal transduction [2] and cleavage of hydrophobic anchors from membrane proteins [3].

A specific and potent inhibitor of PLC could be a useful pharmacological tool, but to date, the inhibitors found among natural substances are not useful in therapy [4]. Syntheses of analogues of natural *myo*-inositol phosphates and phosphatidyl-*myo*-inositol have been reported, but none of them is a potent and specific inhibitor of PI-PLC [5].

Phosphonates have been used previously in the study of lipases [6] and, recently, the preparation [7] and the effects on the activity [8] of several bacterial PI-specific phospholipase C (PI-PLC) of phosphonate derivatives of *myo*-inositol were reported.

We now describe the synthesis and the inhibitory activity on PI-PLC from human platelets of two acylphosphonates of *myo*-inositol.

* Corresponding author.



Scheme 1.

Dimethyl acylphosphonates were formed by adding dropwise, at room temperature, an equimolar amount of trimethyl phosphite to the corresponding acyl chlorides **1a** and **1b** with yields of 90–95%, respectively. The resulting acylphosphonates **2a** and **2b** were transesterified with trimethylsilyl bromide in dry dichloromethane and, after evaporation of the solvent at reduced pressure, the products were coupled with (\pm)-1,2,4,5,6-penta-*O*-benzyl-*myo*-inositol (PBI) [10] in dry pyridine and triethylamine in the presence of 2,4,6-triisopropylbenzensulfonyl chloride (TPSCI). The products **3a** and **3b** were isolated by gradient elution on a column of silica gel in yields of 21–22%, respectively, and debenzylated using BF_3 -etherate in ethanethiol [11] (yields 70–43%, respectively) to give **4a** and **4b**.

The inhibitory activity of **4a** and **4b** on PI-PLC from human platelets was measured under conditions previously described [12]. The two compounds, at a concentration of 10^{-4} M, gave 50% inhibition.

Assuming that the acylphosphonic bond cannot be hydrolysed by PI-PLC, compounds **4a** and **4b** can be assumed to be true inhibitors thus differing from octadecylphosphodithionyl-1-*myo*-inositol [12] and a deoxy analogue of phosphatidylinositol [13]. The former, prepared by us within this program, was found to have similar activity, while the latter was found to be a twenty times less potent inhibitor of PI-PLC than **4a** and **4b** [13].

1. Experimental

General methods.—Melting points were determined with a Reichert Thermovar apparatus and are uncorrected. TLC and column chromatography were performed on silica gel (E. Merck).

The ^1H NMR spectra were recorded at 25°C using a Bruker (200 MHz) spectrometer with the appropriate internal standard. IR spectra were recorded for solutions in CHCl_3 using a Mattson 3000 FTIR spectrometer.

Dimethyl octadecanoylphosphonate (2a).—Trimethyl phosphite (0.7 mL, 6 mmol) was added dropwise to octadecanoyl chloride **1a** (2.0 mL, 6 mmol) under N_2 . After 3 h at room temperature, TLC (CHCl_3) showed the absence of **1a**. The mixture was concentrated under vacuum to give a waxy product **2a** (2.26 g, 95%) which was used without further purification; ν_{max} 2930 (m), 2857 (m), 1697 (m), 1465 (w), 1260 (m), 1183 (w), 1038 (s), 838 (w) cm^{-1} . ^1H NMR data (200 MHz, CDCl_3): δ 3.85 (d, 6 H, J 10.7 Hz, OCH_3), 2.81 (t, 2 H, J 7.1 Hz, CH_2CO), 1.61 (t, 2 H, J 7.1 Hz, $\text{CH}_2\text{CH}_2\text{CO}$), 1.24 (s, 28 H, $\text{CH}_3(\text{CH}_2)_{14}$), 0.87 (t, 3 H, J 6.5 Hz, CH_3). Anal. Calcd for $\text{C}_{20}\text{H}_{41}\text{O}_4\text{P}$: C, 63.80; H, 10.98; P, 8.23. Found: C, 63.92; H, 11.00; P, 8.38.

Dimethyl hexadecanoylphosphonate (2b).—Using the same conditions as described for **2a**, **1b** (1.82 mL, 6 mmol) was treated with trimethyl phosphite (0.7 mL, 6 mmol) to give the waxy product **2b** (2.05 g, 98%) which was used without further purification; ν_{max} 2925 (m), 2854 (m), 1698 (m), 1464 (w), 1258 (m), 1183 (w), 1038 (s), 840 (w) cm^{-1} . ^1H NMR data (200 MHz, CDCl_3): δ 3.81 (d, 6 H, J 10.7 Hz, OCH_3), 2.77 (t, 2 H, J 7.2 Hz, CH_2CO), 1.59 (t, 2 H, J 7.0 Hz, $\text{CH}_2\text{CH}_2\text{CO}$), 1.22 (s, 24 H, $\text{CH}_3(\text{CH}_2)_{12}$), 0.85 (t, 3 H, J 6.5 Hz, CH_3). Anal. Calcd for $\text{C}_{18}\text{H}_{37}\text{O}_4\text{P}$: C, 62.04; H, 10.70; P, 8.89. Found: C, 62.02; H, 10.66; P, 8.90.

1-O-octadecanoylphosphonyl-2,3,4,5,6-penta-O-benzyl-myoinositol (3a).—Bromotrimethylsilane (1.95 mL, 15 mmol) was added dropwise to a solution of **2a** (2.26 g, 5.7 mmol) in dry CH_2Cl_2 (10 mL) under N_2 . After 1 h at room temperature, TLC (65:35:5 CHCl_3 – CH_3OH – NH_4OH) showed the complete disappearance of **2a** (R_f 0.9). The solution was concentrated and the oily residue treated with dry pyridine (25 mL) and dry triethylamine (4.8 mL). Then (±)-1,2,4,5,6-penta-O-benzyl-myoinositol (3.29 g, 5.22 mmol) and 2,4,6-triisopropylbenzenesulfonyl chloride (5.7 g, 18.8 mmol) were added, and the mixture was stirred at room temperature under N_2 . After 24 h, water (2 mL) was added to the mixture and, after concentration under reduced pressure, dry Et_2O was added. The resulting precipitate was filtered and the filtrate was concentrated to give an oily dark red residue which was chromatographed on SiO_2 eluting with CHCl_3 and 98:2 and 95:5 CHCl_3 – CH_3OH to give **3a** as a syrup (1.88 g, 21%). ^1H NMR data (200 MHz, CDCl_3): δ 7.26 (m, 25 H, 5 Ph), 4.62 (m, 12 H, 5 CH_2Ph + H-1 + H-2), 4.35 (t, 1 H, J 10 Hz, H-6), 4.20 (t, 1 H, J 10 Hz, H-4), 3.53 (m, 2 H, H-5 + H-3), 2.85 (t, 2 H, J 10 Hz, CH_2CO), 1.26 (s, 30 H, $\text{CH}_3(\text{CH}_2)_{15}$), 0.86 (t, 3 H, J 10 Hz CH_3). Anal. Calcd for $\text{C}_{66}\text{H}_{84}\text{O}_9\text{P}$: C, 75.33; H, 8.05; P, 2.94. Found: C, 75.31; H, 8.06; P, 2.90.

1-O-hexadecanoylphosphonyl-2,3,4,5,6-penta-O-benzyl-myoinositol (3b).—Using the conditions described for **3a**, **2b** (0.992 g, 2.85 mmol) was treated with bromotrimethylsilane (0.98 mL, 7.5 mmol) and then with (±)-1,2,4,5,6-penta-O-benzyl-myoinositol (1.645 g, 2.61 mmol) and 2,4,6-triisopropylbenzenesulfonyl chloride (2.85 g, 9.4 mmol) to give **3b**

as a syrup (504 mg, 0.54 mmol; 22%). ^1H NMR data (200 MHz, CDCl_3): δ 7.28 (m, 25 H, 5 Ph), 4.75 (m, 12 H, 5 CH_2Ph + H-1 + H-2), 4.17 (t, 1 H, J 10 Hz, H-6), 3.78 (m, 1 H, H-4), 3.42 (m, 2 H, H-5 + H-3), 2.70 (t, 2 H, J 12 Hz, CH_2CO), 1.23 (s, 26 H, $\text{CH}_3(\text{CH}_2)_{13}$), 0.85 (t, 3 H, J 12 Hz, CH_3). Anal. Calcd for $\text{C}_{64}\text{H}_{80}\text{O}_9\text{P}$: C, 75.05; H, 7.87; P, 3.02. Found: C, 75.02; H, 7.86; P, 3.00.

1-O-octadecanoylphosphonyl-myo-inositol (4a).—To a solution of **3a** (340 mg, 0.354 mmol) in dry CH_2Cl_2 (7 mL) ethanethiol (6.86 mL) and boron trifluoride etherate (1.4 mL) were added dropwise at room temperature under N_2 . After 1 h, water was added and the aqueous phase was washed repeatedly with Et_2O and lyophilized to give a solid residue which was purified by chromatography using 65:35:5 CHCl_3 – CH_3OH – H_2O as eluent to give **4a** (126 mg, 0.247 mmol; 70%) as a white solid; mp 244–246°C (from CH_3OH – H_2O); ν_{max} 3360 (s, br), 2922 (s), 2850 (s), 1750 (w), 1189 (m), 1042 (m) cm^{-1} . ^1H NMR data (200 MHz, CDCl_3): δ 4.57–4.44 (m, 5 H, 5 OH), 3.92 (t, 1 H, J 3 Hz, H-2), 3.82 (m, 1 H, H-1), 3.45 (m, 2 H, H-6 + H-4), 2.74 (m, 2 H, CH_2CO), 1.59 (s, 30 H, $\text{CH}_3(\text{CH}_2)_{15}$), 0.87 (t, 3 H, J 10 Hz CH_3). Anal. Calcd for $\text{C}_{24}\text{H}_{47}\text{O}_9\text{P}$: C, 56.45; H, 9.28; P, 6.07. Found: C, 56.44; H, 9.31; P, 6.11.

1-O-hexadecanoylphosphonyl-myo-inositol (4b).—Using the conditions described for **4a**, **3b** (439 mg, 0.53 mmol) was treated with ethanethiol (10.2 mL) and boron trifluoride etherate (2.1 mL) to give **4b** as a white solid, further crystallised from CH_3OH – H_2O to give 110 mg (0.23 mmol, 43%) of **4b**; mp 236–238°C. ν_{max} 3360 (s, br), 2920 (s), 2852 (s), 1748 (w), 1184 (m), 1036 (m) cm^{-1} . ^1H NMR data (200 MHz, CDCl_3): δ 4.49 (m, 5 H, 5 OH), 3.93 (t, 1 H, J 2 Hz, H-2), 3.67–3.52 (m, 2 H, H-1 + H-6), 3.36 (m, 1 H, H-4), 3.10–2.88 (m, 4 H, H-5 + H-3 + CH_2CO), 1.24 (s, 26 H, $\text{CH}_3(\text{CH}_2)_{13}$), 0.85 (t, 3 H, J 8 Hz, CH_3). Anal. Calcd for $\text{C}_{22}\text{H}_{43}\text{O}_9\text{P}$: C, 54.76; H, 8.98; P, 6.42. Found: C, 54.80; H, 9.01; P, 6.40.

References

- [1] R.H. Michell, *Trends Biochem. Sci.*, (1992) 274; M.J. Berridge and R.F. Irvine, *Nature (London)*, 341 (1989) 197–205; A.A. Abdel-Latif, *Pharmacol. Rev.*, 38 (1986) 227–272.
- [2] M.J. Berridge, *Nature (London)*, 361 (1993) 315–325; C.P. Downes, *Biochem. Soc. Trans.*, 17 (1989) 259–268.
- [3] T.L. Doering, W.J. Masterson, G.W. Hart, and P.T. Englund, *J. Biol. Chem.*, 265 (1990) 611–614.
- [4] H. Ogawara, K. Higashi, S. Manita, K. Tanaka, Y. Shimizu, and L. Shufang, *J. Antibiot.*, 45 (1992) 1365–1366; M. Aoki, Y. Itezono, H. Shirai, N. Nakayama, A. Sakai, Y. Tanaka, A. Yamaguchi, N. Simma, K. Yokose, and H. Seto, *Tetrahedron Lett.*, 32 (1991) 4737–4740; T. Nishikiori, A. Okuyama, H. Naganawa, T. Takita, M. Hamada, T. Takeuchi, T. Aoyagi, and H. Umezawa, *J. Antibiot.*, 37 (1984) 426–427.
- [5] A.B. Reitz (Ed.), *Inositol Phosphates and Derivatives, Synthesis, Biochemistry, and Therapeutic Potential*, ACS Symp. Ser., 463 (1991).
- [6] W. Yuan and M.H. Gelb, *J. Am. Chem. Soc.*, 110 (1988) 2665–2666.
- [7] S.S. Yang, T.R. Beattie, P.L. Durette, T.F. Gallagher, and T.-Y. Shen, U.S. Patent No. 4,515,722 (1985); M.S. Shashidhar, J.F.W. Keana, J.J. Volwerk, and O. Hayes Griffith, *Chem. Phys. Lipids*, 53 (1990) 103–113.
- [8] M.S. Shashidhar, J.J. Volwerk, J.F.W. Keana, and O. Hayes Griffith, *Biochim. Biophys. Acta*, 1042 (1990) 410–412.
- [9] G.M. Kosolapoff and L. Maier, *Organic Phosphorus Compounds*, Vol. 5, Wiley, New York, 1976.
- [10] R. Gigg and C.D. Warren, *J. Chem. Soc. C*, (1969) 2367–2371.

- [11] K. Fuji, K. Ichikawa, M. Node, and E. Fuijta, *J. Org. Chem.*, 44 (1979) 1661–1664.
- [12] M.A. Alisi, M. Brufani, L. Filocamo, G. Gostoli, L. Cellai, M.A. Iannelli, G. Melino, M.C. Cesta, and S. Lappa, *Bioorg. Med. Chem. Lett.*, 3 (1993) 1931–1934.
- [13] S.P. Seitz, R.F. Kaltenbach III, R.H. Vreekamp, J.C. Calabrese, and F.W. Perrella, *Bioorg. Med. Chem. Lett.*, 2 (1992) 171–174.